

<https://doi.org/10.17221/628/2020-PSE>

## Effects of hydrogen peroxide application on agronomic traits of rice (*Oryza sativa* L.) under drought stress

WEERAPHORN JIRA-ANUNKUL, WATTANA PATTANAGUL\*

Department of Biology, Faculty of Science, Khon Kaen University, Khon Kaen, Thailand

\*Corresponding author: [wattana@biology.in.th](mailto:wattana@biology.in.th)

**Citation:** Jira-anunkul W., Pattanagul W. (2021): Effects of hydrogen peroxide application on agronomic traits of rice (*Oryza sativa* L.) under drought stress. Plant Soil Environ., 67: 221–229.

**Abstract:** Drought stress is a major environmental factor limiting crop growth and productivity. Hydrogen peroxide ( $H_2O_2$ ) plays an essential role during stress response by acting as a signal molecule that activates multiple stress tolerance mechanisms. In this study, the effects of  $H_2O_2$  on agronomic traits were studied in rice (*Oryza sativa* L.) cv. Khao Dawk Mali 105 (KDML 105) was subjected to drought stress.  $H_2O_2$  was applied by either seed priming or foliar application method with a concentration of 1, 5, and 15 mmol/L. The results showed that both seed priming and foliar application with  $H_2O_2$  improved some yield components. The tiller numbers, number of panicles, number of filled grains, filled grain weight, and harvest index were improved approximately 1.13, 1.04, 1.23, 1.21, and 1.1 times compared to the untreated plants. Foliar application, however, helps the plant by reducing yield loss as indicated by a 0.5-time reduction in the number of unfilled grain and lower unfilled grain weight. It was suggested that 5 mmol/L  $H_2O_2$  was the most effective concentration to alleviate the effect of drought stress during the reproductive stage in rice.

**Keywords:** abiotic stress; drought tolerance; foliar spray; rainfall; antioxidant enzyme; stress acclimation

Drought is considered one of the most severe environmental factors limiting crop growth and productivity. Rice cultivation in many areas of Thailand still largely depends on rainfall, and drought has become a significant problem affecting its yield. The previous study suggested that drought strongly influences rice growth and development, particularly in the reproductive stage (Singh et al. 2018). Zhang et al. (2018) conducted a meta-analysis of 55 published studies and found that drought decreased the agronomic traits by 25.4% among varying growth stages. Besides, rice yield decreased by 53–92% and 48–94% during exposure to mild and severe drought stress (Lafitte et al. 2007). For example, Yang et al. (2019) reported that rice physiological traits and yield decreased by 23.2–24% during the flowering stage. Approximately 67% of Thailand's total rice-growing area is accounted for rain-fed cultivation, especially in the northeastern region of Thailand, and the lowest average grain yield was reported in this area (Jongdee et al. 2006, Haefele and Konboon 2009).

Under unfavourable conditions, plants produce excess amount reactive oxygen species (ROS), which causes peroxidation of lipids, denaturation of proteins, DNA mutation, disruption of cellular homeostasis, and various types of cellular oxidative damage (Pandey and Shukla 2015). Among ROS, hydrogen peroxide ( $H_2O_2$ ) also plays an essential role in many developmental stages such as germination, growth, senescence, and signaling response to stresses (Černý et al. 2018).  $H_2O_2$  has been used to improve plants' tolerance against various environmental stresses. Seed priming and foliar spray methods are standard methods for  $H_2O_2$  application due to their simplicity and inexpensiveness. Using  $H_2O_2$  as a priming agent in wheat resulted in increased growth, water potential, osmolytes accumulation, antioxidant system, and higher yield under drought stress (Habib et al. 2020). Similarly, foliar application with low concentrations of  $H_2O_2$  helped protect yield loss in cotton by increasing antioxidant enzyme activities (Sarwar

Supported by the Development and Promotion of Science and Technology Talents Project (DPST) and Salt-tolerant Rice Research Group, Khon Kaen University, Thailand.

et al. 2017). The beneficial effects of  $H_2O_2$  were also reported in wheat in which 1 000-grain weight, grain per spike, and some physiological traits were improved under drought stress (Farooq et al. 2017).

Most studies on the effects of  $H_2O_2$  application were performed in the vegetative stage; only a few studies have focused on the reproductive stage. In addition, no data has been reported on the comparison between seed priming and foliar application method under drought stress. Therefore, it is interesting to determine the effects of  $H_2O_2$  on agronomic traits in rice and compare the results between seed priming and foliar application method. The knowledge gained may improve rice productivity by alleviating the effect of drought stress during the reproductive stage.

## MATERIAL AND METHODS

**Plant materials and growing conditions.** Rice cultivation was performed during August–November 2019 at the greenhouse at the Department of Biology, Faculty of Science, Khon Kaen University, Khon Kaen, Thailand. The average humidity during August to November 2019 was 72.45, 74.66, 65.95, and 62.56%, respectively. The average temperature during August to November 2019 was 29.72, 29.07, 29.59, and 27.23 °C, respectively. The experimental design was carried out using a completely randomised design (CRD) with five replicates in each treatment. The experiment was divided into two groups of  $H_2O_2$  application methods, including seed priming and foliar spray. For seed priming treatment, 200 rice seeds were fully immersed in 200 mL in priming solutions, including 1 mmol/L  $H_2O_2$  (SP1), 5 mmol/L  $H_2O_2$  (SP5), and 15 mmol/L  $H_2O_2$  (SP15) at a temperature of 25 °C for 24 h in dark condition. For control, untreated plants, and foliar spray application, the seeds were fully immersed in distilled water. The treated and untreated seeds were then germinated and sown in 20.32 cm plastic pot containing 4.5 kg soil (loamy sand; pH 6.04; organic matter 0.24%; electrical conductivity 0.04 dS/m; total nitrogen 0.02%; total phosphorus 36.5 mg/kg; total potassium 232.5 mg/kg and cation exchange capacity 3.94 cmol<sub>+</sub>/kg). Two rice seeds were sown in each pot and grown until the plant reached the developmental stage of phase 14–15 on the BBCH scale (21 days). After that, one vigor seedling was selected, while the other was removed and continued to grow until BBCH 45–47 phase (70 days). Before subjected to drought stress, the plants in the foliar spray group were treated with different

concentrations of  $H_2O_2$ , including 1 mmol/L (FP1), 5 mmol/L (FP5), and 15 mmol/L (FP15). In contrast, the control and untreated groups were sprayed with distilled water. The plants were sprayed (20 mL/spray/pot) to a runoff in early morning (8.00 a.m.) and repeated for two days. On day 72 of cultivation, the plants were all exposed to drought stress by reducing irrigation by 25% of the field capacity every two weeks except the control group, which was watered daily. Rice yield was recorded at 120 days after planting.

**Plant growth and yield component measurements.** Plant height was recorded at day 30 (BBCH 14–15), 60 (BBCH 29–30), 72 (BBCH 45–47), and day 120 (BBCH 99) after planting. For biomass measurement, the plants were separated into the root and the shoot parts. Shoot length, root length and dry weight (DW) of shoot and root were recorded. For dry weight, the samples were dried in a hot air oven at 80 °C for three days, and the weight of each sample was recorded.

At harvesting, yield components were recorded, including the number of tillers/hill and the number of panicles/hill. Panicle length was taken from the base of the rachis to the tip of each panicle. The total number of (filled and unfilled) grains/panicle and hill were manually counted. Filled and unfilled grains were determined by floating the seeds in the water (the submerged grains defined as filled grains) (Zhang et al. 2015). Filled and unfilled weight per panicle, 1 000-grain weight were measured. The harvest index was calculated according to the following formula: harvest index (%) = grain yield × 100/biological yield (Farooq et al. 2017).

**Statistical analysis.** Statistical analysis using one-way ANOVA and all means were separated at the  $P < 0.05$  level using Duncan's multiple range test. Pearson correlation coefficients were calculated to determine the relationship among all parameters.

## RESULTS

**Growth parameters.** In this study, plants on days 30, 60, 72 were not subjected to drought stress, but the data is also reported. The results showed that seed priming with  $H_2O_2$  significantly increased plant height during both unstressed and drought conditions. On day 30, the data showed that seed priming with 1 and 5 mmol/L  $H_2O_2$  (SP1 and SP5) significantly increased plant height by 11–13% compared to the control and untreated group. On days 60 and 72,

<https://doi.org/10.17221/628/2020-PSE>

seed priming with 5 and 15 mmol/L (SP5 and SP15) also increased plant height by approximately 7–13% compared to the control plant. In the foliar-spray group, all treatments were previously primed with distilled water; therefore, no significant difference was observed during the well-watered period (days 30, 60, and 72) (Table 1).

The plants were subjected to drought stress at day 72 to 120. At day 120, seed priming with all  $H_2O_2$  increased plant height compared to the control and untreated plants, especially when primed with 15 mmol/L  $H_2O_2$ . With this concentration, the plant height was improved by 12.10% and 6.89% compared to the control and untreated groups. Foliar-sprayed with  $H_2O_2$ , however, did not enhance plant height as observed in seed priming treatment. Plant height was slightly reduced when sprayed with 5 and 15 mmol/L  $H_2O_2$  (Table 1).

The dry weight of both root and shoot parts was dramatically decreased after exposure to drought stress. Root dry weight in the untreated group was reduced by 56.93% compared to the control plant, which was watered daily. On the contrary, shoot dry weight was not much affected by drought stress, as its level was not significantly different between the control and untreated group. Treating with  $H_2O_2$ , both seed priming and foliar application, had no effect on root and shoot dry weights compared to the untreated group (Table 1).

**Yield components.** Drought stress significantly reduced yield, as indicated by a 23.26% reduction in the number of tillers/hill in the untreated group compared to the control plants. Application of  $H_2O_2$

with both methods slightly increased the number of tillers/hill on average of 14.14% in all concentrations compared to the untreated group, although the numbers were not significantly different (Table 2). Drought also reduced the number of total panicles/hill by 14.28% when compared to the unstressed plant. While seed priming with  $H_2O_2$  did not positively affect, the number of panicles/hill, foliar-spray with  $H_2O_2$  seems to help by increasing the number of panicles/hill, especially at high concentrations (Table 2).

Panicle length and the number of primary branches/panicle were not affected when the plant was under drought stress. Similarly,  $H_2O_2$  application also had no effect on both the panicle length and the number of primary branches/panicle in all concentrations (Table 2). Harvest index (HI) was calculated from grain yield per dry weight. The results showed that HI significantly decreased by 36.81% in the stressed plant compared to the control plant. Seed priming with 5 and 15 mmol/L  $H_2O_2$  significantly increased HI by 37.27% and 30.67% compared to the untreated plant, as well as foliar spray with 5 mmol/L  $H_2O_2$ , which increased by 39.40% (Table 2).

When the rice plant was subjected to drought stress, the number of filled grains/panicle and the number of filled grains/hill were significantly reduced by 35.77% and 51.45% compared to control, respectively. Seed priming with  $H_2O_2$  resulted in a higher number of filled grains/panicle and the number of filled grains/hill, especially those primed with 5 mmol/L  $H_2O_2$ , which increase the number of filled grains/panicle and the number of filled grains/

Table 1. Growth parameters of rice cv. KDML 105 grown after exposure to drought stress

Treatment	Plant height (cm)				Dry weight (g)	
	day 30	day 60	day 72	day 120	root	shoot
Control	84.02 ± 5.05 <sup>a</sup>	120.88 ± 2.53 <sup>a</sup>	153.98 ± 2.52 <sup>ab</sup>	174.32 ± 2.54 <sup>abc</sup>	93.80 ± 12.60 <sup>c</sup>	36.94 ± 0.97 <sup>b</sup>
Untreated	82.44 ± 2.55 <sup>a</sup>	130.14 ± 3.07 <sup>ab</sup>	163.36 ± 8.76 <sup>abc</sup>	182.82 ± 2.89 <sup>bcd</sup>	40.40 ± 7.60 <sup>ab</sup>	34.23 ± 0.71 <sup>ab</sup>
SP1	93.58 ± 1.35 <sup>b</sup>	127.72 ± 4.17 <sup>ab</sup>	167.54 ± 5.80 <sup>bc</sup>	188.16 ± 4.59 <sup>cd</sup>	47.75 ± 5.00 <sup>ab</sup>	35.87 ± 0.64 <sup>ab</sup>
SP5	93.34 ± 1.82 <sup>b</sup>	130.16 ± 5.68 <sup>ab</sup>	174.52 ± 6.74 <sup>c</sup>	186.86 ± 5.83 <sup>cd</sup>	32.88 ± 2.42 <sup>ab</sup>	33.13 ± 1.15 <sup>ab</sup>
SP15	89.04 ± 1.45 <sup>ab</sup>	135.18 ± 1.71 <sup>b</sup>	175.06 ± 4.85 <sup>c</sup>	195.42 ± 6.19 <sup>d</sup>	25.56 ± 2.85 <sup>a</sup>	32.26 ± 1.07 <sup>ab</sup>
FP1	89.04 ± 2.33 <sup>ab</sup>	136.18 ± 1.92 <sup>b</sup>	172.76 ± 3.63 <sup>bc</sup>	184.16 ± 2.22 <sup>bcd</sup>	37.03 ± 2.56 <sup>ab</sup>	31.69 ± 2.71 <sup>a</sup>
FP5	94.24 ± 1.93 <sup>b</sup>	128.68 ± 5.14 <sup>ab</sup>	154.90 ± 8.23 <sup>ab</sup>	169.74 ± 6.83 <sup>ab</sup>	39.95 ± 5.00 <sup>ab</sup>	32.31 ± 0.99 <sup>ab</sup>
FP15	89.98 ± 3.86 <sup>ab</sup>	124.42 ± 3.96 <sup>ab</sup>	148.12 ± 5.33 <sup>a</sup>	162.22 ± 5.22 <sup>a</sup>	48.92 ± 11.68 <sup>b</sup>	32.64 ± 2.13 <sup>ab</sup>

The plants were watered daily from day 0 to day 72 and were exposed to drought stress from day 72 to day 120. Data (means ± standard error,  $n = 5$ ) followed with different letters within the same column indicate a significant difference according to Duncan's multiple range test ( $P < 0.05$ ). SP1 – 1 mmol/L seed priming; SP5 – 5 mmol/L seed priming; SP15 – 15 mmol/L seed priming; FP1 – 1 mmol/L foliar spray; FP5 – 5 mmol/L foliar spray; FP15 – 15 mmol/L foliar spray

Table 2. Yield component of rice cv. KDML 105 grown after exposed to drought stress

Treatment	No. of tillers/hill	No. of total panicle/hill	Panicle length/panicle (cm)	Number of primary branch/panicle	Harvest index (%)
Control	8.60 ± 0.40 <sup>b</sup>	9.80 ± 0.20 <sup>bc</sup>	34.88 ± 0.32 <sup>ab</sup>	10.74 ± 0.37 <sup>a</sup>	31.89 ± 0.60 <sup>e</sup>
Untreated	6.60 ± 0.40 <sup>a</sup>	8.40 ± 0.68 <sup>ab</sup>	34.99 ± 0.71 <sup>ab</sup>	11.46 ± 0.20 <sup>ab</sup>	20.15 ± 1.52 <sup>ab</sup>
SP1	8.20 ± 0.20 <sup>ab</sup>	8.60 ± 0.40 <sup>abc</sup>	35.39 ± 0.67 <sup>ab</sup>	11.15 ± 0.14 <sup>ab</sup>	21.47 ± 1.65 <sup>abc</sup>
SP5	7.60 ± 0.68 <sup>ab</sup>	7.80 ± 0.66 <sup>a</sup>	36.68 ± 0.33 <sup>b</sup>	11.24 ± 0.24 <sup>ab</sup>	27.66 ± 2.54 <sup>cde</sup>
SP15	7.20 ± 0.20 <sup>ab</sup>	7.80 ± 0.37 <sup>a</sup>	35.17 ± 0.60 <sup>ab</sup>	11.08 ± 0.17 <sup>ab</sup>	26.33 ± 1.96 <sup>bcde</sup>
FP1	6.80 ± 0.58 <sup>ab</sup>	8.80 ± 0.37 <sup>abc</sup>	35.96 ± 0.43 <sup>ab</sup>	11.47 ± 0.13 <sup>ab</sup>	24.95 ± 2.55 <sup>abcd</sup>
FP5	8.00 ± 0.84 <sup>ab</sup>	9.40 ± 0.60 <sup>bc</sup>	35.84 ± 1.11 <sup>ab</sup>	11.72 ± 0.44 <sup>b</sup>	28.09 ± 1.78 <sup>de</sup>
FP15	7.40 ± 0.75 <sup>ab</sup>	10.00 ± 0.32 <sup>c</sup>	33.86 ± 0.57 <sup>a</sup>	11.29 ± 0.24 <sup>ab</sup>	19.44 ± 2.94 <sup>a</sup>

Data (means ± standard error,  $n = 5$ ) followed with different letters within the same column indicate a significant difference according to Duncan's multiple range test ( $P < 0.05$ ). SP1 – 1 mmol/L seed priming; SP5 – 5 mmol/L seed priming; SP15 – 15 mmol/L seed priming; FP1 – 1 mmol/L foliar spray; FP5 – 5 mmol/L foliar spray; FP15 – 15 mmol/L foliar spray

hill by 31.58% and 50.11%, respectively. Foliar spray with 5 mmol/L  $H_2O_2$  also improved the number of filled grains/panicle by 28.28% and 50.29% in the number of filled grain/hill compared to the untreated plant (Figure 1A, B).

On the contrary, drought stress caused significant increases in the number of unfilled grains/panicle by 56.61% and a slight increase in the number of unfilled grains/hill by 17.56%. While seed priming with  $H_2O_2$  alleviated the effect of drought by decreasing the number of unfilled grains/panicle and the number of unfilled grains/hill, the result was more prominent in the plant foliar-sprayed with  $H_2O_2$  at all concentrations. Foliar application with  $H_2O_2$  decreased the number of unfilled grains/panicle by 48–57%, as well as the number of unfilled grains/hill, which was reduced by 38–51% (Figure 1C, D).

Filled grain weight/panicle and filled grain weight/hill were dramatically decreased by 37.10% and 52.27%, respectively, when the rice plant was subjected to drought stress. Seed priming and foliar-spray with  $H_2O_2$  slightly increased the filled grain weight/panicle in all concentrations compared to the untreated plant, except those sprayed with 15 mmol/L  $H_2O_2$ . In comparison, both seed priming and foliar spray with 5 mmol/L  $H_2O_2$  resulted in the highest filled grain weight/panicle. In addition, filled grain weight/hill was also improved when the plants were treated with  $H_2O_2$ . Seed priming and foliar-sprayed with 5 mmol/L  $H_2O_2$  resulted in a significantly higher filled grain weight/hill than the untreated plant (Figure 1E, F).

Drought stress also caused increases in the unfilled grain weight/panicle and the unfilled grain weight/

hill by 79.49% and 34.23% compared to the untreated plant. Seed priming with  $H_2O_2$  increased plant yield by decreasing the unfilled grain weight/panicle and the unfilled grain weight/hill. Also, foliar-spray with  $H_2O_2$  was even more effective than the seed priming method since the unfilled grain weight/panicle, and the unfilled grain weight/hill were lower than those in the untreated group and the seed priming treatment (Figure 1G, F).

One-thousand-grain weight was significantly reduced by 24.74% when the plant was subjected to drought stress. Seed priming with 1 and 5 mmol/L  $H_2O_2$ , as well as a foliar spray with 5 mmol/L  $H_2O_2$ , slightly improved 1 000-grain weight by 22.63, 19.87, and 17.67%, respectively (Figure 1I).

Pearson's correlation coefficient among the results was presented in Table 3. It was found that plant height was not positively correlated with yield-related traits, while root dry weight (RDW) was positively correlated with shoot dry weight (SDW), the total number of panicles (TP), number of filled grain/hill (NFH), and filled grain weight/hill (FWH). Shoot dry weight was positively correlated with the number of tillers (NT), number of unfilled grains/hill (NUH), and 1 000-grain weight (TWH). For yield-related components, harvest index was positively correlated with panicle length (PL), number of filled grains/hill, filled grain weight/hill, number of filled grains/panicle (NFP), and filled grain weight/panicle (FWP). The number of filled grains/hill was positively correlated with root dry weight, the number of tillers, harvest index, filled grain weight/hill, 1 000-grain weight, and filled grain weight/panicle.

<https://doi.org/10.17221/628/2020-PSE>

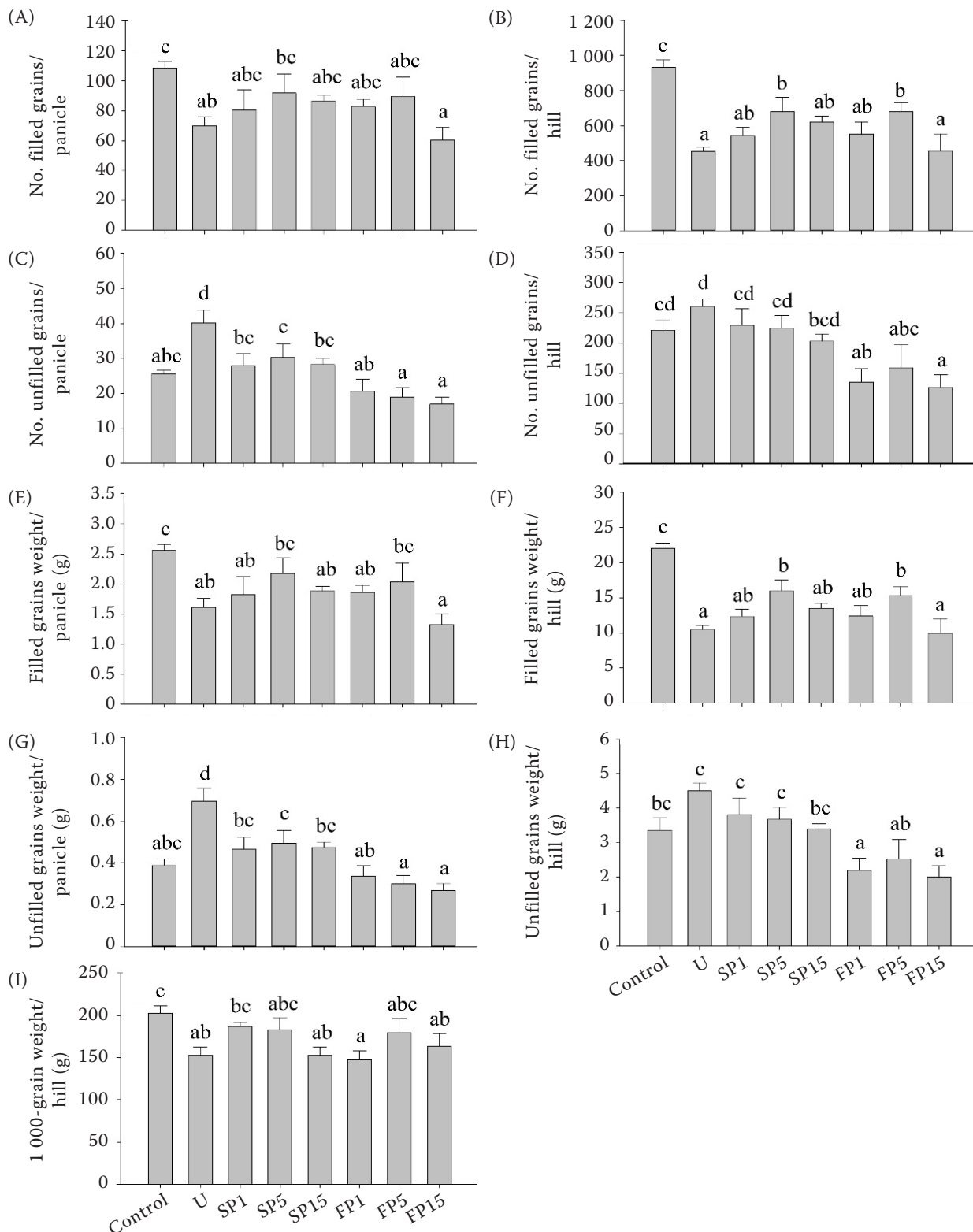


Figure 1. The effects of hydrogen peroxide ( $H_2O_2$ ) on yield components of rice cv. KDML 105 under drought stress. Data are shown as the mean  $\pm$  standard error ( $n = 5$ ). (A) Number of filled grains/panicle; (B) number of filled grains/hill; (C) number of unfilled grains/panicle; (D) number of unfilled grains/hill; (E) filled grain weight/panicle; (F) filled grain weight/hill; (G) unfilled grain weight/panicle; (H) unfilled grain weight/hill, and (I) 1 000-grain weight. U – untreated; SP1 – 1 mmol/L seed priming; SP5 – 5 mmol/L seed priming; SP15 – 15 mmol/L seed priming; FP1 – 1 mmol/L foliar spray; FP5 – 5 mmol/L foliar spray; FP15 – 15 mmol/L foliar spray



Table 3. Pearson's correlation between parameters in drought-stressed rice

	H120	RDW	SDW	NT	TP	HI	PL	NB	NFH	NUH	FWH	UWH	TWH	NFP	NUP	FWP	UWP
H120		-0.310	-0.128	-0.315*	-0.646**	0.258	0.391*	-0.012	0.146	0.203	0.144	0.239	-0.337*	0.376*	0.378*	0.366*	0.391*
RDW			0.481**	0.284	0.446**	0.225	-0.166	-0.137	0.423**	0.075	0.474**	-0.017	0.354*	0.265	-0.061	0.299	-0.139
SDW				0.485**	0.058	-0.134	0.026	0.058	0.194	0.521**	0.220	0.457**	0.506**	-0.029	0.349*	-0.009	0.285
NT					0.405**	0.190	-0.228	-0.336*	0.409**	0.360*	0.386*	0.298	0.935**	-0.074	-0.137	-0.089	-0.170
TP						-0.051	-0.302	-0.206	0.034	-0.265	0.018	-0.304	0.359*	-0.161	-0.513**	-0.179	-0.525**
HI							0.335*	-0.032	0.902**	-0.138	0.902**	-0.201	0.221	0.807**	-0.244	0.812**	-0.286
PL								0.430**	0.250	-0.181	0.277	-0.203	-0.141	0.480**	-0.060	0.502**	-0.086
NB									-0.094	-0.289	-0.105	-0.219	-0.315*	0.114	-0.051	0.109	0.005
NFH										0.043	0.988**	-0.043	0.410**	0.786**	-0.164	0.772**	-0.223
NUH											0.070	0.965**	0.416**	-0.210	0.860**	-0.177	0.813**
FWH												-0.022	0.432**	0.783**	-0.125	0.795**	-0.194
UWH													0.356*	-0.260	0.858**	-0.233	0.874**
TWH														-0.056	-0.057	-0.019	-0.092
NFP															-0.171	0.984**	-0.209
NUP																-0.130	0.970**
FWP																	-0.176
UWP																	

\*Correlation is significant at the 0.05 level (2-tailed); \*\*Correlation is significant at the 0.01 level (2-tailed). H120 – plant height at 120 days; RDW – root dry weight; SDW – shoot dry weight; NT – no. of tillers; TP – total no. panicles; HI – harvest index; PL – panicle length; NB – no. of the primary branch; NFH – no. filled grains/hill; NUH – no. unfilled grains/hill; FWH – filled grain weight/hill; UWH – unfilled weight/hill; TWH – 1 000 grain weight/hill; NFP – no. filled grains/panicle; NUP – no. unfilled grains/panicle; FWP – filled grains weight/panicle; UWP – unfilled grains weight/panicle

<https://doi.org/10.17221/628/2020-PSE>

## DISCUSSIONS

In plants,  $H_2O_2$  plays diverse roles in various biochemical and physiological processes. Due to its long life span and versatility,  $H_2O_2$  can traverse through cellular membranes and potentially act as a signaling molecule in the stress signal transduction pathway. These pathways can then trigger various responses involved in stress acclimation. This study showed that seed priming with appropriate concentrations of  $H_2O_2$ , i.e., 1 mmol/L and 5 mmol/L, enhanced plant growth under non-stressed as well as drought conditions. In cannabis, using  $H_2O_2$  as a priming agent showed an increase in dry weight under normal conditions (Golizadeh et al. 2015). Also, seed priming with  $H_2O_2$  shows higher dry weight and root length under drought conditions in wheat (Hameed and Iqbal 2014).

It was proposed that seed priming could leave the "stress memory," which, in turn, activates the stress response mechanisms after germination (Chen and Arora 2013). Seed priming with a high concentration of  $H_2O_2$ , however, resulted in the reduction of root and shoot biomass. Despite acting as a signaling molecule, a high concentration of  $H_2O_2$  may lead to oxidative stress that causes cellular damage and results in programmed cell death (Gupta et al. 2016).

Yield components, including tiller number, harvest index, panicle length, as well as number and weight of filled grains, were also improved by  $H_2O_2$  priming. Priming with 5 mmol/L  $H_2O_2$  was the most effective for improving rice yield. It has been hypothesised that seed priming could activate stress-responsive systems such as antioxidant activation, hormonal regulation, and abscisic acid signaling (Chen and Arora 2013, Lutts et al. 2016). Cheng et al. (2017) reported that the genes associated with stress were observed at the early imbibition stage. Besides, genes related to protein synthesis, carbohydrate metabolism, and signaling were up-regulated at the late imbibition stage. Marthandan et al. (2020) proposed that the seed priming method can induce physiological adaptation, which in turn increases crop yield under drought stress. In the review, the stress memory left in the seed can faster, and stronger activate the gene and transcription after post-germination. In wheat, the yield was also enhanced under drought stress when primed with an appropriate concentration of  $H_2O_2$  (Hameed and Iqbal 2014). Habib et al. (2020) reported that  $H_2O_2$  priming has the potential to increase tiller, 1 000-grain weight, and grain yield/plant. Optimal concentrations of  $H_2O_2$  can enhance abiotic stress

tolerance through the modulation of multiple physiological processes, such as photosynthesis, stomatal movement, osmotic adjustment, and ROS detoxification (Quan et al. 2008, Niu and Liao 2016, Sayed and Gadallah 2019). In mustard, seed priming with  $H_2O_2$  showed significant increases in antioxidant enzyme activities (Hossain and Fujita 2013). This result indicates that  $H_2O_2$  could trigger the ROS detoxification mechanism, which is crucial for maintaining the structural and membrane integrity of cellular organelles and keeping them fully functional in plants under abiotic stresses (Hossain et al. 2015). The study in *Caklie martima* Scop. showed that seed priming with 120  $\mu$ mol  $H_2O_2$  improved malondialdehyde content (MDA), lipid peroxidation index in both drought and salt stress (Ellouzi et al. 2017). A recent study in rice also found that seed priming with 5 mmol/L  $H_2O_2$  decreased electrolyte leakage but increased relative water content (Jira-anunkul and Pattanagul 2020).

Although foliar-spray with  $H_2O_2$  did not affect shoot growth and biomass during drought stress, it is interesting that spraying with  $H_2O_2$  did increase root growth in which may increase the plant's capability to access water. Yield components, on the other hand, were significantly improved by foliar application with  $H_2O_2$ . The number of filled grain-filled grain weight and harvest index were higher than the untreated plant, especially in 5 mmol/L treatment. Rice reproductive stage at the flowering stage is highly susceptible to drought leading to pollen and spikelet sterility, which causes a significant reduction in yield (Yang et al. 2019). There was evidence suggested that  $H_2O_2$  application helps increase yield in many plants. Orabi et al. (2018) reported that spraying with 2 mmol  $H_2O_2$  induced the antioxidant enzyme activities, e.g., catalase, ascorbate peroxidase, while decreased lipid peroxidation in the canola plant. The onion yield was significantly improved when applied with 1–2 mmol/L  $H_2O_2$  (Semida 2016). Therefore, it is suggested that physiological changes caused by  $H_2O_2$  help the plants maintain growth under unfavourable conditions, which results in a better yield.

In addition, it is noteworthy that foliar-spray with  $H_2O_2$  significantly lower the unfilled grain number and weight, which was more prominent than the seed priming method. A previous study suggested that  $H_2O_2$  might activate the defense system as well as improve water status in rice under drought stress. It is also reported that  $H_2O_2$  treatment decreased the endogenous  $H_2O_2$ , which resulted in lower malondialdehyde content.  $H_2O_2$  accumulation also triggers

kinase protein and targets calcium homeostasis, ion channel, phosphatase, transcription factor, and ABA signaling response to stress (Černý et al. 2018). The previous evidence suggested that pretreatment with  $H_2O_2$  increased abscisic acid and osmolytes concentration in maize leaves (Terzi et al. 2014). Absciscic acid is well-documented for triggering stomatal closure and activating stress-responsive gene expression (Ali et al. 2020). The antioxidant system, in particular, is triggered by  $H_2O_2$  signaling and may account for the observed increase in total yield as well as preventing yield loss. Taken all together, it is suggested that  $H_2O_2$  application prevents the plant from ROS attack and maintains normal physiological processes, therefore, prevent yield loss (Sohag et al. 2020). Similar to the seed priming treatment, a high concentration of  $H_2O_2$  does not enhance yield production but instead results in yield loss.

In conclusion, both seed priming and foliar spray with  $H_2O_2$  contributed to improving rice cv. KDML 105 growth and agronomic traits under drought stress. Using 5 mmol  $H_2O_2$  as a priming agent or foliar application was the most effective concentration as indicated by increases in filled grain weight, filled grain number, and harvest index. Similar to the seed priming method, foliar application with  $H_2O_2$  provided benefits in increasing yield production and also preventing yield loss. Therefore, the decision on each technique depends on available facilities, cost, and farmer's preference.

**Acknowledgment.** We would like to thank the Department of Biology, Faculty of Science, Khon Kaen University for providing tools and facilities and Khon Kaen Rice Research Center for supplying seeds in this experiment.

## REFERENCES

- Ali S., Hayat K., Iqbal A., Xie L.N. (2020): Implications of abscisic acid in the drought stress tolerance of plants. *Agronomy*, 10: 1323.
- Černý M., Habánová H., Berka M., Luklová M., Brzobohatý B. (2018): Hydrogen peroxide: its role in plant biology and crosstalk with signalling networks. *International Journal of Molecular Sciences*, 19: 2812.
- Chen K.T., Arora R. (2013): Priming memory invokes seed stress-tolerance. *Environmental and Experimental Botany*, 94: 33–45.
- Cheng J., Wang L., Zeng P., He Y., Zhou R., Zhang H., Wang Z. (2017): Identification of genes involved in rice seed priming in the early imbibition stage. *Plant Biology*, 19: 61–69.
- Ellouzi H., Sghayar S., Abdelly C. (2017):  $H_2O_2$  seed priming improves tolerance to salinity; drought and their combined effect more than mannitol in *Cakile maritima* when compared to *Eutrema salsugineum*. *Journal of Plant Physiology*, 210: 38–50.
- Farooq M., Nawaz A., Chaudhary M.A.M., Rehman A. (2017): Foliage-applied sodium nitroprusside and hydrogen peroxide improves resistance against terminal drought in bread wheat. *Journal of Agronomy and Crop Science*, 203: 473–482.
- Golizadeh S.K., Mahmoodi T.M., Khaliliaqdam N. (2015): The effects of priming by folic acid and hydrogen peroxide on morpho-physiological traits in cannabis seeds (*Cannabis sativa* L.). *Journal of Biodiversity and Environmental Sciences*, 6: 407–416.
- Gupta K., Sengupta A., Chakraborty M., Gupta B. (2016): Hydrogen peroxide and polyamines act as double edged swords in plant abiotic stress responses. *Frontiers in Plant Science*, 7: 1343.
- Habib N., Ali Q., Ali S., Javed M.T., Haider M.Z., Perveen R., Shahid M.R., Rizwan M., Abdel-Daim M.M., Elkelish A., Bin-Jumah M. (2020): Use of nitric oxide and hydrogen peroxide for better yield of wheat (*Triticum aestivum* L.) under water deficit conditions: growth, osmoregulation, and antioxidative defense mechanism. *Plants (Basel)*, 9: 285.
- Haefele S.M., Konboon Y. (2009): Nutrient management for rainfed lowland rice in northeast Thailand. *Field Crops Research*, 114: 374–385.
- Hameed A., Iqbal N. (2014): Chemo-priming with mannose, mannitol and  $H_2O_2$  mitigate drought stress in wheat. *Cereal Research Communications*, 42: 450–462.
- Hossain M.A., Bhattacharjee S., Armin S.-M., Qian P.P., Xin W., Li H.Y., Burritt D.J., Fujita M., Tran L.S.P. (2015): Hydrogen peroxide priming modulates abiotic oxidative stress tolerance: insights from ROS detoxification and scavenging. *Frontiers in Plant Science*, 6: 420.
- Hossain M.A., Fujita M. (2013): Hydrogen peroxide priming stimulates drought tolerance in mustard (*Brassica juncea* L.) seedlings. *Plant Gene and Trait*, 4: 109–123.
- Jira-anunkul W., Pattanagul W. (2020): Seed priming with hydrogen peroxide alleviates the effects of drought stress in rice (*Oryza sativa* L.) seedlings. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 48: 273–283.
- Jongdee B., Pantuwan G., Fukai S., Fischer K. (2006): Improving drought tolerance in rainfed lowland rice: an example from Thailand. *Agricultural Water Management*, 80: 225–240.
- Lafitte H.R., Yongsheng G., Yan S., Li Z.K. (2007): Whole plant responses, key processes, and adaptation to drought stress: the case of rice. *Journal of Experimental Botany*, 58: 169–175.
- Lutts S., Benincasa P., Wojtyła L., Kubala S.S., Pace R., Lechowska K., Quinet M., Garnczarska M. (2016): Seed priming: new comprehensive approaches for an old empirical technique. In: Araújo S. (ed.): *New Challenges in Seed Biology – Basic and Translational Research Driving Seed Technology*. Rijeka, IntechOpen, 1–47. ISBN: 978-953-51-2659-1



<https://doi.org/10.17221/628/2020-PSE>

- Marthandan V., Greetha R., Kumutha K., Ranganathan V.G., Karthikeyan A., Ramalingam J. (2020): Seed priming: a feasible strategy to enhance drought tolerance in crop plants. *International Journal of Molecular Sciences*, 21: 8258.
- Niu L.J., Liao W.B. (2016): Hydrogen peroxide signaling in plant development and abiotic responses: crosstalk with nitric oxide and calcium. *Frontiers in Plant Science*, 7: 230.
- Orabi S.A., Hussein M.M., Szaki S., Sharara F.A. (2018): Influence of hydrogen peroxide on growth, yield and biochemical constituents of canola plants grown under different irrigation intervals. *Current Science International*, 7: 407–418.
- Pandey V., Shukla A. (2015): Acclimation and tolerance strategies of rice under drought stress. *Rice Science*, 22: 147–161.
- Quan L.J., Zhang B., Shi W.W., Li H.Y. (2008): Hydrogen peroxide in plants: a versatile molecule of the reactive oxygen species network. *Journal of Integrative Plant Biology*, 50: 2–18.
- Sarwar M., Saleem M.F., Najeeb U., Shakeel A., Ali S., Bilal M.F. (2017): Hydrogen peroxide reduces heat-induced yield losses in cotton (*Gossypium hirsutum* L.) by protecting cellular membrane damage. *Journal of Agronomy and Crop Science*, 203: 429–441.
- Sayed S., Gadallah M. (2019): Hydrogen peroxide supplementation alleviates the deleterious effects of cadmium on photosynthetic pigments and oxidative stress and improves growth, yield and pods quality of pea (*Pisum sativum* L.) plants. *Acta Physiologiae Plantarum*, 41: 113.
- Semida W.M. (2016): Hydrogen peroxide alleviates salt-stress in two onion (*Allium cepa* L.) cultivars. *American-Eurasian Journal of Agricultural and Environmental Sciences*, 16: 294–301.
- Singh S., Prasad S., Yadav V., Kumar A., Jaiswal B., Kumar A., Khan N.A., Dwivedi D.K. (2018): Effect of drought stress on yield and yield components of rice (*Oryza sativa* L.) genotypes. *International Journal of Current Microbiology and Applied Sciences*, 7: 2752–2759.
- Sohag A.A.M., Tahjib-Ul-Arif M., Brestic M., Afrin S., Sakil M.A., Hossain M.T., Hossain M.A., Hossain M.A. (2020): Exogenous salicylic acid and hydrogen peroxide attenuate drought stress in rice. *Plant, Soil and Environment*, 66: 7–13.
- Terzi R., Kadioğlu A., Kalaycıoğlu E., Sağlam A. (2014): Hydrogen peroxide pretreatment induces osmotic stress tolerance by influencing osmolyte and abscisic acid levels in maize leaves. *Journal of Plant Interactions*, 9: 559–565.
- Yang X.L., Wang B.F., Chen L., Li P., Cao C.G. (2019): The different influences of drought stress at the flowering stage on rice physiological traits, grain yield, and quality. *Scientific Reports*, 9: 3742.
- Zhang J.M., Zhang S.Q., Cheng M., Jiang H., Zhang X.Y., Peng C.H., Lu X.H., Zhang M.X., Jin J.X. (2018): Effect of drought on agronomic traits of rice and wheat: a meta-analysis. *International Journal of Environmental Research and Public Health*, 15: 839.
- Zhang Y.J., Wang Z.S., Li L., Zhou Q., Xiao Y., Wei X., Zhou M.Y. (2015): Short-term complete submergence of rice at the tillering stage increases yield. *PLoS One*, 10: e0127982.

Received: December 3, 2020

Accepted: March 3, 2021

Published online: March 23, 2021